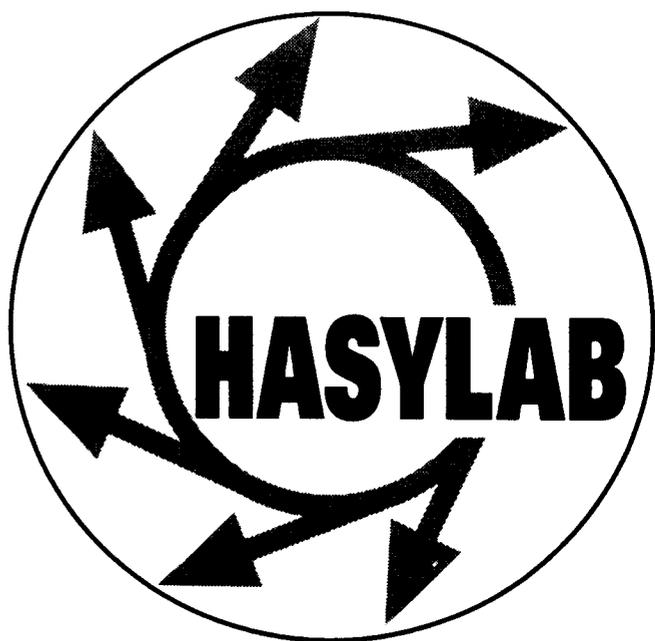


# Jahresbericht 1993

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Hamburger Synchrotronstrahlungslabor  
**HASYLAB**  
am Deutschen Elektronen-Synchrotron  
**DESY**



## X-ray small angle scattering of Cytochrome-c Oxidase (cytochrome-aa<sub>3</sub>) and Cytochrome-c Reductase (cytochrome-bc<sub>1</sub>) from beef heart mitochondria

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Cytochrome-c reductase (cytochrome-bc<sub>1</sub> complex) and cytochrome-c oxidase (cytochrome-aa<sub>3</sub> complex) are large membrane protein complexes of energy conserving systems in many organisms, e.g. man [1]. These enzymes couple a redox reaction to the transport of protons across the membrane and thus the biological oxidation to the formation of an electrochemical proton potential difference of the membrane. These biological redox-catalysts consist of a functional core, which is present also in bacterial proteins, and additional protein subunits which are found in cytochrome proteins of higher organisms only.

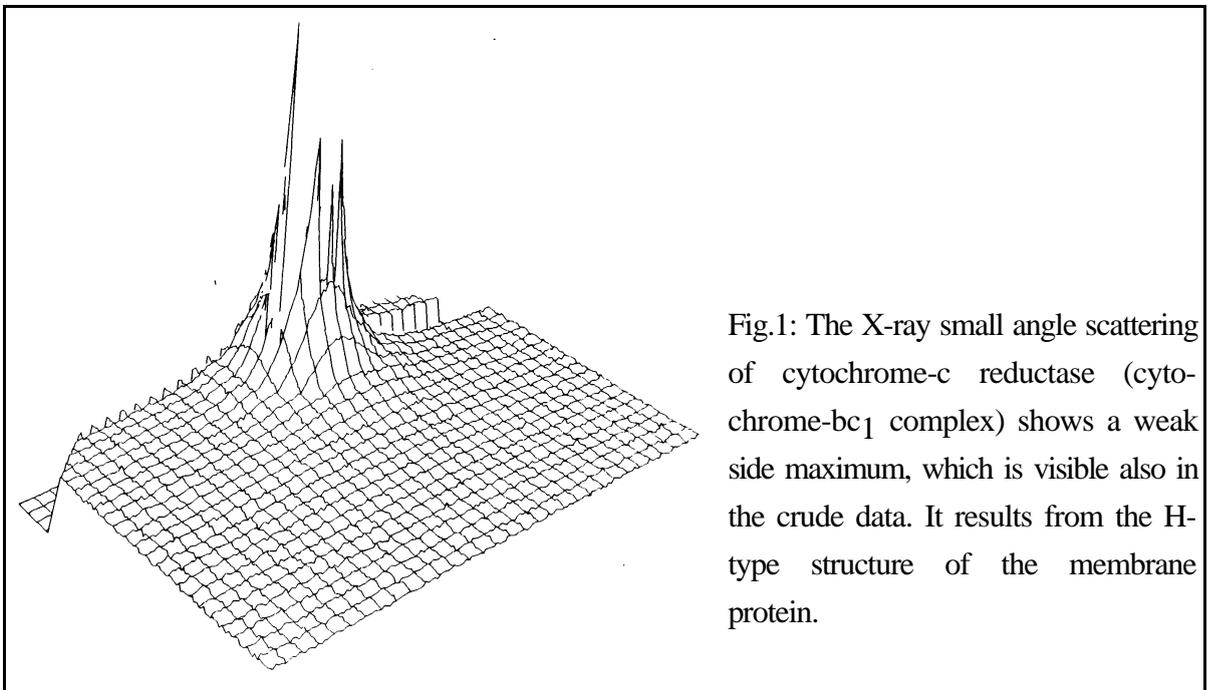
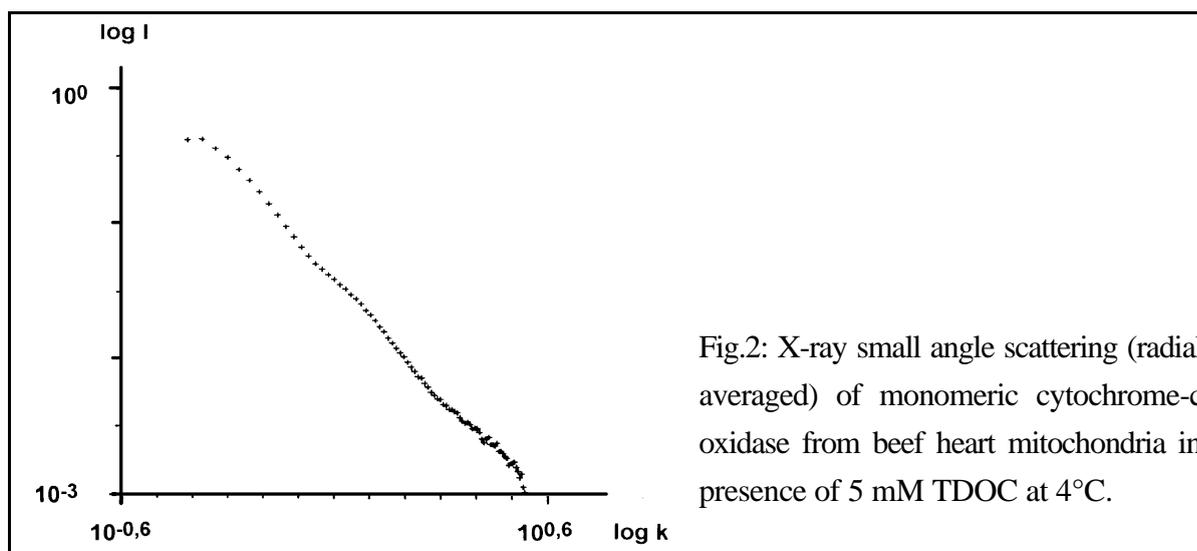


Fig.1: The X-ray small angle scattering of cytochrome-c reductase (cytochrome-bc<sub>1</sub> complex) shows a weak side maximum, which is visible also in the crude data. It results from the H-type structure of the membrane protein.

We have isolated, purified and characterized cytochrome-c reductase and cytochrome-c oxidase from beef heart mitochondria [2,3]. By detergent solubilization studies and analytical ultracentrifugation we found that functional cytochrome-c reductase is always a dimer,

whereas the dimeric cytochrome-c oxidase can be split into monomeric native enzyme entities. The structure of the enzymes was investigated at the A1 beamline at HASYLAB/DESY using radiation of 0.15 nm wavelength. The detector-sample distance was 1.9 m. The detergent contribution to the over all scattering of cytochrome-c reductase was small because of the small surfactant concentration (0.1% w/v laurylmaltoside) and the nearly contrast-matching buffer. In case of cytochrome-c oxidase we solubilized the enzyme by 5 mM taurodeoxycholate (TDOC), which was earlier shown to yield only negligible scattering because of its very small mizelle size [4]. The residual micelle scattering was eliminated by subtracting the scattering of a protein-free detergent buffer.

The X-ray small angle scattering of cytochrome-c reductase showed a side maximum, that results of the H-type structure of the dimeric enzyme (fig.1) [5]. The small angle scattering of monomeric cytochrome-c oxidase in solution of 5 mM TDOC (fig.2) showed only a less pronounced shoulder. This corresponds to the proposed structure of a long flat asymmetric 'y'. The radius of gyration of this enzyme was  $R_g = 3.73$  nm. The scattering of both enzymes is now interpreted by molecular models consisting of cubes according to the FVM-method [6].



- 1) Saraste, M. (1990) *Quart.Rev.Biophys.* 23, 331-366
- 2) Link, T.A.; Schägger, H.; von Jagow, G. (1986) *FEBS Lett.* 204, 9-15
- 3) Gencic, S.; Schägger, H.; von Jagow, G. (1991) *Eur. J. Biochem.* 199, 123-131
- 4) Conrad, H.; Dose, K.; Nawroth, T. (1989) *Physica* 156 B, 474-476
- 5) Bechmann, G.; Schulte, U.; Weiss, H. (1992) in: *Molecular Mechanisms in Bioenergetics* (ed.: L.Ernster) *New Comprehensive Biochemistry*, Vol.23, Elsevier, Amsterdam, pp. 199-216
- 6) Nawroth, T. (1989) *Physica* 156 B, 493-495