



EXPERIMENTAL REPORT

EXPERIMENT N° 8-03-302

INSTRUMENT D22

DATES OF EXPERIMENT 29. 09. 1998

TITLE **In situ structure of membrane bound ATP-synthase from *Micrococcus luteus* in contrast-matched liposomes**

EXPERIMENTAL TEAM (names and affiliation)

T. Nawroth, I. Lauer : Biochemistry Institute, Gutenberg-University, Becherweg 30, D-55099 Mainz

Tel.: 0 049 6131 395702; <http://www.uni-mainz.de/FB/Chemie/Biochemie/MPSD/TNa.html>

J. Holzinger, F. Descamps, R.P. May : ILL, Grenoble

LOCAL CONTACT **Roland P. May**

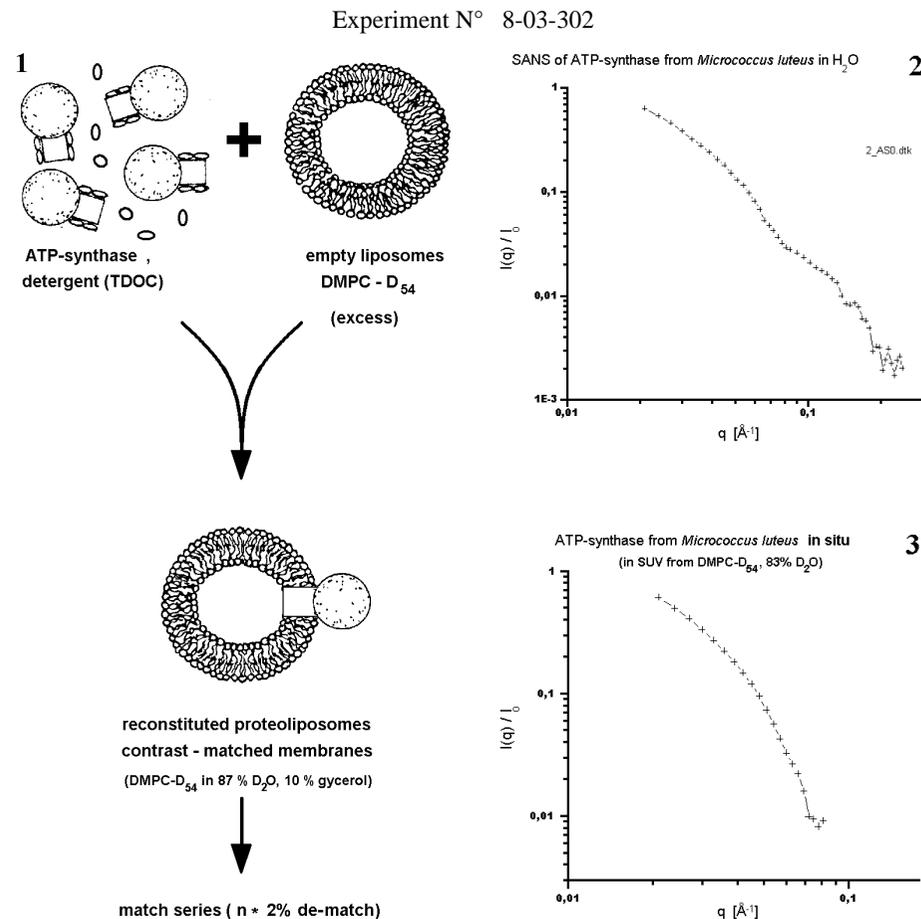
Date of report 15.3.1999

The structure of membrane proteins in situ, i.e. bound to membranes, can be exclusively studied by neutron small angle scattering of contrast matched reconstituted proteoliposomes [1,2]. In this study we were successful in improving the method by application to the remarkable stable ATP-synthase from *Micrococcus luteus* in DMPC-D₅₄ liposomes and match variation series (systematic de-matching in 2% steps).

In the experiments three short subexperiment series with each of the particle types in the reconstitution process were done: As shown in fig.1 the proteoliposomes were formed by reconstitution of preformed liposomes from DMPC-D₅₄, ATP-synthase and detergent solution by our liposome - detergent incubation method [1]. The very small contribution of 5 mM TDOC detergent micelles [3] was eliminated by subtracting detergent reference buffer, which yielded the SANS of free ATP-synthase in H₂O (fig.2). The Guinier approximation ($q = 0.021-0.027 \text{ \AA}^{-1}$) yielded a radius of gyration of $R_G = 55.21 \pm 0.13 \text{ \AA}$ (in D₂O: $62.3 \pm 0.5 \text{ \AA}$). The semi-direct Fourier transformation (extrapolation of the missing regions only [by Guinier $q < 0.021 \text{ \AA}^{-1}$; by Porod $0.25 < q < 0.6 \text{ \AA}^{-1}$], then discrete sinus FT) yielded the maximum dimension $r_{\text{max}} = 180 \pm 2 \text{ \AA}$. By contrast variation (0, 68, 97% D₂O content in 10% H-glycerol buffer) the protein scattering length density $\rho = 2.24 \pm 0.03 \cdot 10^{-10} \text{ cm}^{-2}$ was obtained, which is required for the evaluation of the proteoliposome subexperiment.

Pure (protein-free) liposomes (sonicated SUV) from DMPC, SBL and DMPC-D₅₄ were investigated. With SUV from DMPC and SBL time resolved SANS was done as reported in the TEST-261 and TEST-252 experiments. As important improvement we were successful in testing the novel ILL-time-frame controller, which allowed TR-SANS without time-gaps.

With ATP-synthase/DMDC-D₅₄ liposomes and pure reference vesicles match variation series were done by systematic de-matching of slightly over-matched SUV (87% D₂O, 10% H-glycerol). The detergent content (1.67 mM) was not removed for SANS because it stabilizes the liposomes [4]. The subtraction of the scattering at the lipid match point yielded the SANS of ATP-synthase from *Micrococcus luteus* in situ (fig.3): $R_G = 60.64$



Figures:

1) Proteoliposome reconstitution by incubation of preformed DMPC-D₅₄ liposomes (41 g/l), ATP-synthase (4.74 g/l) and detergent (TDOC; finally 1.67 mM = 1/2 cmc); 2) SANS of 3.74g/l ATP-synthase in 5 mM TDOC buffer pH8 (H₂O; 1h); 3) SANS of ATP-synthase of *Micrococcus luteus* (1.58 g/l) in contrast-matched liposomes, i.e. in situ (t = 1h). The valid range will be increased by a factor of 2 by subtraction of match-extrapolated datasets instead of direct experiment files after completion of the KINEX2 evaluation program.

References 1) Nawroth, T.; Conrad, H.; Vienken, J.; Dose, K. (1983) Hoppe Seyler's Z. Physiol. Chemie 364, 923-931
2) Nawroth, T.; Dose, K.; Conrad, H. (1989) Physica B 156 & 157, 489-492
3) Conrad, H.; Dose, K.; Nawroth, T. (1989) Physica B 156 & 157, 474-476
4) Nawroth, T.; Conrad, H.; Dose, K. (1989) Physica B 156 & 157, 477-480